DOI: 10.1002/open.201900110



Conjugates of Cryptophycin and RGD or *iso*DGR Peptidomimetics for Targeted Drug Delivery

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RGD-cryptophycin and *iso*DGR-cryptophycin conjugates were synthetized by combining peptidomimetic integrin ligands and cryptophycin, a highly potent tubulin-binding antimitotic agent across lysosomally cleavable Val-Ala or uncleavable linkers. The conjugates were able to effectively inhibit binding of biotiny-lated vitronectin to integrin $\alpha_{\rm v}\beta_{\rm 3}$, showing a binding affinity in the same range as that of the free ligands. The antiproliferative activity of the novel conjugates was evaluated on human melanoma cells M21 and M21-L with different expression levels of integrin $\alpha_{\rm v}\beta_{\rm 3}$, showing nanomolar potency of all four

compounds against both cell lines. Conjugates containing uncleavable linker show reduced activity compared to the corresponding cleavable conjugates, indicating efficient intracellular drug release in the case of cryptophycin-based SMDCs. However, no significant correlation between the in vitro biological activity of the conjugates and the integrin $\alpha_{\rm v}\beta_3$ expression level was observed, which is presumably due to a non-integrin-mediated uptake. This reveals the complexity of effective and selective $\alpha_{\rm v}\beta_3$ integrin-mediated drug delivery.

1. Introduction

Chemotherapeutic agents used in traditional tumor therapy do not preferentially accumulate at the tumor site and are often associated with poor pharmacokinetics leading to inferior efficacy and severe side effects due to systemic toxicity. An emerging approach to minimize chemotherapy side effects and maximize tumor specificity, is the covalent conjugation of cytotoxic drugs to antibodies that provide site-specific targeted delivery of the toxin. [1–3] The resulting antibody-drug conjugates (ADCs) have a definitive impact on anticancer drug develop-

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Supporting information for this article is available on the WWW under https://doi.org/10.1002/open.201900110

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ment with currently four ADCs on the market (Adcetris®, Kadcyla®, Mylotarg®, and Besponsa®) and approximately 70 candidates in clinical trials. [4] Although ADCs show great clinical benefit and success on the market, further development is required to address limitations like high manufacturing costs, inefficient tissue penetration and immunogenicity, in particular for the treatment of solid tumors. [5] As an alternative, small-molecule-drug conjugates (SMDCs) have the potential to overcome drawbacks of ADCs. In this case, the cytotoxic agent is connected to a small synthetic ligand that selectively binds to an appropriate target overexpressed on the tumor cell surface. [6,7]

Integrins, a family of 24 transmembrane heterodimers composed of 18α and 8β integrin subunits, are considerable targets in anticancer therapy due to their involvement in many steps of cancer progression.^[8] The cell adhesion protein, integrin $\alpha_v \beta_3$ is highly expressed on endothelial and tumor cells of various solid tumors such as glioma, gastric cancer, non-small cell lung cancer, prostate, and pancreatic cancer. [9-11] In addition, integrin $\alpha_v \beta_3$ plays a vital role in cancer progression, angiogenesis, and metastasis. Therefore, it has been widely considered as potential target antigen $^{[8,12,13]}$ Integrin $\alpha_{\nu}\beta_{3}$ binds endogenous ligands, like vitronectin, by recognizing the tripeptide sequence Arg-Gly-Asp (RGD) as the minimal binding motif.[14] As a result, various RGD-containing synthetic peptides and peptidomimetics have been developed and some of them demonstrate high binding affinity to different integrins. [15-17] Several ligands with strong binding affinity and integrin subtype selectivity have been successfully conjugated to different cytotoxic agents, and have been investigated for tumor targeting applications.[18,19]





Figure 1. Chemical structure of ligands: cyclo[DKP-RGD] (1), $NH_2CH_2-cyclo[DKP-RGD]$ (2), cyclo[DKP-isoDGR] (3), $NH_2CH_2-cyclo[DKP-isoDGR]$ (4) and cytotoxic drugs: cryptophycin-52 (5), cryptophycin-55 (6) and cryptophycin-55 glycinate (7).

Besides the RGD sequence, the isoaspartyl-containing sequence isoAsp—Gly—Arg (isoDGR) was identified as an integrin ligand. Hence, there has been increasing interest in employing isoDGR peptides as RGD analogues for integrin targeting. [20,21] There is a growing body of evidence suggesting that this class of ligands are true integrin $\alpha_{\mbox{\tiny V}}\beta_{\mbox{\tiny 3}}$ antagonists, unlike the RGD peptides, which induce integrin conformational changes upon binding, as well as receptor activation. [22,23] Based on these findings, a panel of cyclic RGD and isoDGR peptidomimetic integrin ligands containing a diketopiperazine (DKP) scaffold have been developed. Importantly, the cyclo[DKP-RGD] 1 and cyclo[DKP-isoDGR] **3** (Figure 1) are selective integrin $\alpha_v \beta_3$ ligands with low nanomolar affinity. [24,25] Functionalized analogues 2 and 4 have been conjugated to different anticancer drugs, such as paclitaxel, [26-31] camptothecin [32] and α -amanitin [33] across different linkage systems including ester, ether, carbonate or carbamate bond, involving non-cleavable, disulfide, elastase cleavable NPV, and lysosomally cleavable Val-Ala, Phe-Lys or GFLG linkers.

Although the ligand plays a key role for efficient targeting, the high potency of the cytotoxic drug is also a crucial criterion for optimal SMDC design. Cryptophycins are compelling antimitotic payloads for tumor targeting applications, owing to their high activity as tubulin polymerization inhibitors, relative hydrophilicity and ability to overcome multidrug resistance. Initially the synthetic analogue cryptophycin-52 (5, Figure 1) was developed and clinically investigated as an antitumor agent on its own. However, neurotoxic side effects and the lack of activity led to the failure of this drug candidate in clinical phase II. Sissis Further research has been focused on the investigation of

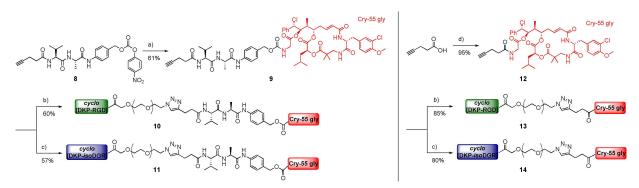
structural moieties that are essential for the biological activity. Consequently, a large variety of synthetic analogues have been designed and tested for structure-activity relationship (SAR) studies.^[37–39] More recently, cryptophycin derivatives bearing a functional handle suitable for attachment to tumor targeting peptides or antibodies have come to the forefront.^[34,40–43] In this context, the unit A-modified chlorohydrin derivative (6, Figure 1) and the glycinate of the chlorohydrin hydroxyl group (7, Figure 1) are highly potent in vitro and in vivo.^[44] The latter has been successfully conjugated to trastuzumab targeting HER2,^[45] acetazolamide targeting CAIX,^[46] and other RGD-based integrin ligands.^[47]

2. Results and Discussion

Here we focus on the conjugation of the cryptophycin-55 glycinate derivative to the functionalized *cyclo*[DKP-RGD] and *cyclo*[DKP-*iso*DGR] peptidomimetics for integrin targeted delivery. The conjugates were designed to connect the integrin ligands to the cytotoxin across the lysosomally cleavable Val—Ala linker (10 and 11, Scheme 1). The cathepsin B sensitive peptide linker combined with the *p*-aminobenzylcarbamate (PABC) self-immolative spacer is widely used to ensure efficient release of the free drug upon endocytosis. Conjugates with uncleavable linker were also prepared (13 and 14, Scheme 1). Presumably the latter are not substrates of enzymatic cleavage; therefore, the drug should be released only after hydrolysis of ligand-linker components resulting in decreased in vitro antitumor activity of the conjugate compared to the unconjugated







Scheme 1. Synthesis of cyclo[DKP-RGD]-PEG4-Val—Ala-PABC-Cry-55gly (10), cyclo[DKP-isoDGR]-PEG4-Val—Ala-PABC-Cry-55gly (11), cyclo[DKP-RGD]-PEG4-uncl-Cry-55gly (13), and cyclo[DKP-isoDGR]-PEG4-uncl-Cry-55gly (14). Reagents and conditions: a) 7, DIPEA, DMF, RT, 4 h; b) cyclo[DKP-RGD]-PEG4-N₃, $cusO_4 \cdot 5H_2O$, sodium ascorbate, 1:1 DMF/H₂O, 35 °C, 24 h; c) cyclo[DKP-isoDGR]-PEG4-N₃, $cusO_4 \cdot 5H_2O$, sodium ascorbate, 1:1 DMF/H₂O, 35 °C, 24 h; d) 7, PyBOP, HOBt, DIPEA, DMF, RT, 4 h.

drug and the conjugates with cleavable linker. To improve the water solubility of the conjugate and ensure flexibility of the construct, a PEG4 spacer was inserted between the ligand and the triazole ring, used as connection point with the linker.

We evaluated the four conjugates for their ability to inhibit binding of biotinylated vitronectin to isolated integrin $\alpha_v\beta_3$ in comparison with the free ligands. We tested the in vitro cytotoxic activity of the conjugates against the human melanoma cells M21 and M21-L with different integrin expression levels.

2.1. Synthesis

The RGD and *iso*DGR peptidomimetic-cryptophycin conjugates **10**, **11**, **13**, and **14** were synthetized as shown in Scheme 1.

The linker Fmoc—Val—Ala—PAB^[27] was deprotected by piperidine and the crude free amine was subsequently reacted with 4-pentynoic acid. Then, the hydroxyl group was activated using bis(4-nitrophenyl) carbonate (Scheme S1 in the Supporting Information). The resulting alkyne-functionalized activated linker 8 was conjugated to cryptophycin-55 glycinate^[46] by forming a carbamate to give compound 9. The PEG4-containing azido-functionalized *cyclo*[DKP-RGD] and *cyclo*[DKP-*iso*DGR] ligands were prepared as previously reported. [29,33] Final conjugation was achieved by copper(I)-catalyzed azide—alkyne cycloaddition (CuAAC "click" reaction) between compound 9 and *cyclo*[DKP-RGD]-PEG4-azide, respectively.

For the synthesis of uncleavable conjugates 4-pentynoic acid was coupled to cryptophycin-55 glycinate (7) *via* amide bond to afford compound 12. This was subjected to CuAAC reaction with the respective ligands to give the final conjugates 13 and 14. Compounds 8–14 were characterized by NMR, HPLC-MS and HRMS (see Supporting Information).

2.2. Competitive Integrin Binding Assays

Conjugates were evaluated in vitro for their binding affinity to human $\alpha_v\beta_3$ integrin in comparison with the free RGD and

Table 1. Inhibition of biotinylated vitronectin binding to human integrin $\alpha_{\nu}\beta_{3}$. IC₅₀ values were determined as the compound concentration necessary for 50% inhibition of biotinylated vitronectin binding as calculated using the software GraphPad Prism. All values are the average \pm standard deviation of duplicate measurements.

Cmpd	Structure	IC ₅₀ [nM] $\alpha_{v}\beta_{3}$
1	cyclo[DKP-RGD]	4.5 ± 1.1
3	cyclo[DKP-isoDGR]	9.2 ± 1.1
10	cyclo[DKP-RGD]-VA-Cry-55gly	$\textbf{7.2} \pm \textbf{1.5}$
11	cyclo[DKP-isoDGR]-VA-Cry-55gly	5.5 ± 2.8
13	cyclo[DKP-RGD]-Cry-55gly	24.1 ± 1.0
14	cyclo[DKP-isoDGR]-Cry-55gly	10.2 ± 0.4

isoDGR ligands. Competitive binding assays were performed by incubation of immobilized $\alpha_{\nu}\beta_{3}$ integrin with increasing concentrations of test compounds (10⁻⁵-10⁻¹² M) in the presence of the ECM protein, vitronectin (1 μg mL⁻¹). The concentrations causing 50% inhibition of biotinylated vitronectin (IC50) are listed in Table 1. As it is shown, free ligands as well as conjugates efficiently inhibit the biotinylated vitronectin binding to $\alpha_v \beta_3$ integrin with IC₅₀ values in the low nanomolar range. Comparing the binding affinity of the conjugates and ligands we can conclude that conjugates retained high binding affinity. The linker type has only a minimal effect, conjugates with uncleavable linker displayed a slightly reduced affinity, presumably because of the shorter linker between the ligand and the drug. Similar results were reported for analogous conjugates of the cyclo[DKP-RGD] and cyclo[DKP-isoDGR] ligands and paclitaxel or $\alpha\text{-amanitin.}^{\scriptscriptstyle{[29,33]}}$ In addition, high selectivity for $\alpha_{\text{\tiny v}}\beta_3$ integrin was reported for cyclo[DKP-RGD]-paclitaxel conjugates involving various lysosomally cleavable Val-Ala or GFLG linkers.[31] This indicates that conjugation of a drug to the respective ligands does not compromise specificity and we expect that the selectivity of the RGD- and isoDGR-cryptophycin conjugates towards $\alpha_{v}\beta_{3}$ integrin subtype is maintained.

2.3. Cell Viability Assays

The cytotoxicity of the unconjugated drug (7) and cryptophycin-conjugates (10, 11, 13, 14) was assessed for two cell lines





with different expression levels of the integrin. The M21 human melanoma ($\alpha_v \beta_3$ positive) and M21-L ($\alpha_v \beta_3$ negative) model was used to characterize in vitro the tumor-targeting ability of the novel conjugates. [49] The presence of the integrin subunit $\alpha_{\rm v}$ and the integrin heterodimer $\alpha_v \beta_3$ on the cell membrane was confirmed by flow cytometry analysis (Figure S1 in the Supporting Information). The cells were treated with increasing concentrations of the free drug and conjugates 10, 11, 13, 14, respectively, for 2 hours, then the medium was replaced with fresh medium and the incubation was continued for additional 70 hours. The short treatment time aimed to minimalize altered antiproliferative activity caused by incidental extracellular linker cleavage and undesired drug release, as well as intended to resemble more closely the in vivo experimental conditions. The cell viability was measured in an MTT (3-(4,5-dimethylthiazol-2yl)-2,5-diphenyltetrazolium bromide) assay (Figure 2, Table 2).

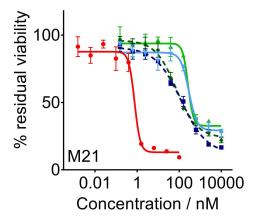
The cryptophycin conjugates display high in vitro potency with IC₅₀ values between 22.3–287.5 nM, showing slightly reduced activity compared to cryptophycin-55 glycinate. Moreover, conjugates bearing cleavable linker (10, 11) are more cytotoxic than the corresponding uncleavable conjugates (13, 14). This underlines that higher potency of the conjugates may be associated with the intracellular cathepsin-mediated drug release, while lower cytotoxicity correlates with less specific linker degradation. Although the intracellular enzymatic cleavage of peptide-linked ADCs and SMDCs is a well-known process,^[50] this study is the first head-to-head comparison for cryptophycin-based SMDCs.

Table 2. Cytotoxic potencies of free cryptophycin-55 glycinate and cryptophycin-conjugates against M21 and M21-L cell lines upon 2 h treatment and 70 h additional incubation

treatment and 70 if additional incubation.			
Structure	$\begin{array}{l} \text{IC}_{50} \ [\text{nM}] \\ \text{M21} \\ (\alpha_{\text{v}}\beta_3 +) \end{array}$	$\begin{array}{l} \text{IC}_{50} [\text{nM}] \\ \text{M21-L} \\ (\alpha_{\text{v}}-,\alpha_{\text{v}}\beta_{3}-) \end{array}$	
Cry-55gly (7) cyclo[DKP-RGD]-VA-Cry-55gly (10) cyclo[DKP-isoDGR]-VA-Cry-55gly (11) cyclo[DKP-RGD]-Cry-55gly (13) cyclo[DKP-isoDGR]-Cry-55gly (14)	0.75 ± 0.11 72.7 ± 11.2 122.6 ± 18.6 261.3 ± 45.8 287.5 ± 35.9	0.14 ± 0.08 22.3 ± 2.0 32.6 ± 3.0 203.0 ± 27.7 140.8 ± 13.3	

All four conjugates were found to exhibit similar cytotoxic activity on the antigen-positive and antigen-negative cell line, indicating that there is no substantial correlation between the expression level of integrin and the observed biological activity. In this regard, it has to be noted that the $\alpha_{\nu}\beta_{3}$ -negative M21-L cell line seems to be more sensitive to cryptophycin and cryptophycin conjugates. It was previously reported that the cytotoxicity of cyclo[DKP-isoDGR]-α-amanitin conjugates was not integrin-specific when tested on human glioblastoma U-87 $(\alpha_v \beta_3 +)$, breast adenocarcinoma MDA-MB-468 $(\alpha_v \beta_3 -)$, and it was hypothesized that the internalization process was mediated by integrins different from $\alpha_v \beta_3$ (e.g. $\alpha_v \beta_5$). This observation prompted us to select the isogenic M21 and M21-L cell lines for the in vitro evaluation of cryptophycin conjugates. The M21-L cell line was considered a valid negative control, as these cells completely lack the integrin α_v subunit. The deficient expression of the α_v at the mRNA level leads to the absence of not only of integrin $\alpha_v \beta_3$, but also the other α_v integrin heterodimers (e.g. $\alpha_v \beta_5$, $\alpha_v \beta_6$). [51] Nevertheless, it is known that the RGD-motif is not exclusively recognized by the α_{v} -integrin subfamily but also by other integrins, namely the $\alpha_{\text{IIb}}\beta_3$, $\alpha_5\beta_1$, and $\alpha_8\beta_1$. For this reason, targeting one distinct integrin subtype using small RGD ligand-based SMDCs is considered to be challenging. Flow cytometry analysis shows similar expression level of integrin $\alpha_5\beta_1$ in the M21 and M21-L cells, that is comparable with the $\alpha_{\nu}\beta_{3}$ integrin expression level in the antigen positive cell line. [53-55] Although cyclic RGD and isoDGR peptidomimetic ligands have been shown to bind to $\alpha_{\scriptscriptstyle 5}\beta_{\scriptscriptstyle 1}$ 100-fold less effective than to $\alpha_v \beta_3$ (1: $\alpha_s \beta_1$ IC₅₀=532 nM,^[56] 3: $\alpha_s \beta_1$ IC₅₀=1066 nM, determined according to ref. 56) the presence of this receptor might play a role in the recognition and internalization of RGDand isoDGR-cryptophycin conjugates, resulting in similar activity against both cell lines.

On the other hand, it has been shown that the *cyclo*-RGDfK-carboxyfluorescein peptide targeting $\alpha_{\rm v}\beta_{\rm 3}$ integrin was taken up by antigen-positive and antigen-negative cells to a similar extent, suggesting that the endocytosis of *cyclo*-RGD peptides is integrin-independent, and proceeds by a fluid-phase pathway. [57] Furthermore, it has been reported that increased



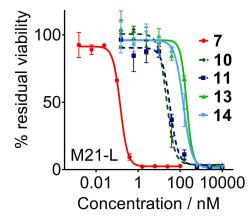


Figure 2. Cell viability determined in an MTT assay with M21 (left), M21-L (right) cells after treatment with Cry-55gly (7), cyclo[DKP-RGD]-cryptophycin and cyclo[DKP-isoDGR]-cryptophycin conjugates. Curves were obtained by non-linear regression, error bars represent the standard deviation of triplicate measurements, the measurements were repeated twice.





hydrophobicity of conjugates caused by the hydrophobic character of the cytotoxic drug may facilitate unwanted non-specific interactions with the cell surface and can result in passive conjugate uptake into the antigen-negative cell.^[58] Indeed, cryptophycins are highly hydrophobic and the cellular uptake of ³H-cryptophycin-52 by THP-1 and H-125 cells was reported to be fast and irreversible.^[59] The nonspecific intracellular uptake of the RGD- and *iso*DGR-cryptophycin conjugates, promoted by the hydrophobic drug or the overall hydrophobicity of the conjugates, may be an additional factor implicated in the observed in vitro potencies. Further increasing the linker hydrophilicity may reduce these undesired interactions and can result in enhanced selectivity.

3. Conclusions

In summary, we have investigated the synthesis and biological activity of novel cyclo[DKP-RGD]-cryptophycin and cyclo[DKPisoDGR]-cryptophycin conjugates aiming at integrin-mediated drug delivery. The cytotoxic payload is connected to the respective peptidomimetic ligand via the protease cleavable Val-Ala linker in combination with PABC self-immolative moiety or a non-cleavable spacer. All four conjugates retain high binding affinity towards isolated $\alpha_v \beta_3$ integrin, similarly to the free ligands. In vitro, the cryptophycin-conjugates were found to be highly potent against both human melanoma cells M21 and M21-L with IC₅₀ values in the nanomolar range. Conjugates containing uncleavable linker demonstrated moderately decreased activity in comparison with analogous cleavable conjugates. Although RGD- and isoDGR-ligands show high specificity for $\alpha_{\mbox{\tiny V}}\beta_{\mbox{\tiny 3}}$ integrin, no correlation could be found between the in vitro antitumor activity of the conjugates and the $\alpha_v \beta_3$ integrin expression level, revealing the complexity of effective and selective $\alpha_v \beta_3$ integrin-mediated drug delivery. We assume that our SMDCs may internalize by an α_v -integrin independent pathway: endocytosis could either be mediated by integrin $\alpha_5\beta_1$, or by nonspecific, fluid-phase uptake, in which the high overall hydrophobicity of the conjugates could also play a role. Nonetheless, the high activity of cryptophycin conjugates encourages us to further investigate and improve the selectivity in tumor-targeted drug delivery.

Acknowledgements

This project has received funding from the European Union's Horizon 2020 research and innovation programme under the Marie Sklodowska-Curie grant agreement No 642004 (ETN MAGICBULLET). We acknowledge the financial support of the German Research Foundation (DFG) and the Open Access Publication Fund of Bielefeld University for the article processing charge. The authors want to acknowledge Georg Falck for flow cytometry analysis, Marco Wißbrock, Anke Nieß, and Carmela Michalek for technical support. Human melanoma cells (M21 and M21-L) were kindly provided by Dr. David Cheresh and The Scripps Research Institute (La Jolla, CA, USA).

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Conflict of Interest

A.M. is an employee of Exiris, P.G. is a shareholder of Exiris. C.S. is a shareholder of Exiris.

Keywords: antitumor agents · cancer · drug delivery integrins · peptidomimetics

- [1] R. V. J. Chari, M. L. Miller, W. C. Widdison, Angew. Chem. Int. Ed. 2014, 53, 3796–3827; Angew. Chem. 2014, 126, 3872–3904.
- [2] A. Beck, L. Goetsch, C. Dumontet, N. Corvaïa, Nat. Rev. Drug Discovery 2017, 16, 315–337.
- [3] J. M. Lambert, C. Q. Morris, Adv. Ther. 2017, 34, 1015-1035.
- [4] Z.-R. Lu, P. Qiao, Mol. Pharm. 2018, 15, 3603-3616.
- [5] N. Krall, J. Scheuermann, D. Neri, Angew. Chem. Int. Ed. 2013, 52, 1384– 1402; Angew. Chem. 2013, 125, 1424–1443.
- [6] C. S. Kue, A. Kamkaew, K. Burgess, L. V. Kiew, L. Y. Chung, H. B. Lee, *Med. Res. Rev.* 2016, 36, 494–575.
- [7] M. Srinivasarao, P. S. Low, Chem. Rev. 2017, 117, 12133–12164.
- [8] H. Hamidi, J. Ivaska, Nat. Rev. Cancer 2018, 18, 533-548.
- [9] M. Nieberler, U. Reuning, F. Reichart, J. Notni, H.-J. Wester, M. Schwaiger, M. Weinmüller, A. Räder, K. Steiger, H. Kessler, Cancers 2017, 9, 116.
- [10] J. S. Desgrosellier, D. A. Cheresh, Nat. Rev. Cancer 2010, 10, 9-22.
- [11] F. Danhier, A. Le Breton, V. Préat, Mol. Pharm. 2012, 9, 2961-2973.
- [12] C. J. Avraamides, B. Garmy-Susini, J. A. Varner, Nat. Rev. Cancer 2008, 8, 604–617.
- [13] H. Jin, J. Varner, Br. J. Cancer 2004, 90, 561-565.
- [14] E. Ruoslahti, M. Pierschbacher, Science 1987, 238, 491-497.
- [15] T. G. Kapp, F. Rechenmacher, S. Neubauer, O. V. Maltsev, E. A. Cavalcanti-Adam, R. Zarka, U. Reuning, J. Notni, H.-J. Wester, C. Mas-Moruno, J. Spatz, B. Geiger, H. Kessler, Sci. Rep. 2017, 7, 39805.
- [16] L. Auzzas, F. Zanardi, L. Battistini, P. Burreddu, P. Carta, G. Rassu, C. Curti, G. Casiraghi, Curr. Med. Chem. 2010, 17, 1255–1299.
- [17] K. Gottschalk, H. Kessler, Angew. Chem. Int. Ed. 2002, 41, 3767–3774; Angew. Chem. 2002, 114, 3919–3927.
- [18] S. Katsamakas, T. Chatzisideri, S. Thysiadis, V. Sarli, Future Med. Chem. 2017, 9, 579–604.
- [19] T. Chatzisideri, G. Leonidis, V. Sarli, Future Med. Chem. 2018, 10, 2201– 2226.
- [20] A. Corti, F. Curnis, J. Cell Sci. 2011, 124, 515-522.
- [21] A. Spitaleri, S. Mari, F. Curnis, C. Traversari, R. Longhi, C. Bordignon, A. Corti, G.-P. Rizzardi, G. Musco, J. Biol. Chem. 2008, 283, 19757–19768.
- [22] M. Ghitti, A. Spitaleri, B. Valentinis, S. Mari, C. Asperti, C. Traversari, G. P. Rizzardi, G. Musco, Angew. Chem. Int. Ed. 2012, 51, 7702–7705; Angew. Chem. 2012, 124, 7822–7825.
- [23] F. Nardelli, C. Paissoni, G. Quilici, A. Gori, C. Traversari, B. Valentinis, A. Sacchi, A. Corti, F. Curnis, M. Ghitti, M. Ghitti, J. Med. Chem. 2018, 61, 7474–7485.
- [24] A. S. M. da Ressurreição, A. Vidu, M. Civera, L. Belvisi, D. Potenza, L. Manzoni, S. Ongeri, C. Gennari, U. Piarulli, Chem. Eur. J. 2009, 15, 12184–12188.
- [25] M. Mingozzi, A. Dal Corso, M. Marchini, I. Guzzetti, M. Civera, U. Piarulli, D. Arosio, L. Belvisi, D. Potenza, L. Pignataro, C. Gennari, *Chem. Eur. J.* 2013, 19, 3563–3567.
- [26] R. Colombo, M. Mingozzi, L. Belvisi, D. Arosio, U. Piarulli, N. Carenini, P. Perego, N. Zaffaroni, M. De Cesare, V. Castiglioni, E. Scanziani, C. Gennari, J. Med. Chem. 2012, 55, 10460–10474.
- [27] A. Dal Corso, M. Caruso, L. Belvisi, D. Arosio, U. Piarulli, C. Albanese, F. Gasparri, A. Marsiglio, F. Sola, S. Troiani, B. Valsasina, L. Pignataro, D. Donati, C. Gennari, Chem. Eur. J. 2015, 21, 6921–6929.
- [28] S. Zanella, S. Angerani, A. Pina, P. López Rivas, C. Giannini, S. Panzeri, D. Arosio, M. Caruso, F. Gasparri, I. Fraietta, C. Albanese, A. Marsiglio, L. Pignataro, L. Belvisi, U. Piarulli, C. Gennari, Chem. Eur. J. 2017, 23, 7910–7914.
- [29] A. Raposo Moreira Dias, A. Pina, A. Dal Corso, D. Arosio, L. Belvisi, L. Pignataro, M. Caruso, C. Gennari, Chem. Eur. J. 2017, 23, 14410–14415.
- [30] A. Raposo Moreira Dias, A. Pina, A. Dean, H.-G. Lerchen, M. Caruso, F. Gasparri, I. Fraietta, S. Troiani, D. Arosio, L. Belvisi, L. Pignataro, A. Dal Corso, C. Gennari, Chem. Eur. J. 2019, 25, 1696–1700.





- [31] P. López Rivas, I. Ranđelović, A. Raposo Moreira Dias, A. Pina, D. Arosio, J. Tóvári, G. Mező, A. Dal Corso, L. Pignataro, C. Gennari, Eur. J. Org. Chem. 2018, 2018, 2902–2909.
- [32] A. Pina, A. Dal Corso, M. Caruso, L. Belvisi, D. Arosio, S. Zanella, F. Gasparri, C. Albanese, U. Cucchi, I. Fraietta, A. Marsiglio, L. Pignataro, D. Donati, C. Gennari, ChemistrySelect 2017, 2, 4759–4766.
- [33] L. Bodero, P. López Rivas, B. Korsak, T. Hechler, A. Pahl, C. Müller, D. Arosio, L. Pignataro, C. Gennari, U. Piarulli, Beilstein J. Org. Chem. 2018, 14, 407–415.
- [34] C. Weiss, E. Figueras, A. N. Borbely, N. Sewald, *J. Pept. Sci.* **2017**, *23*, 514–531.
- [35] G. D'Agostino, J. del Campo, B. Mellado, M. A. Izquierdo, T. Minarik, L. Cirri, L. Marini, J. L. Perez-Gracia, G. Scambia, Int. J. Gynecol. cancer 2006, 16.71–76.
- [36] M. J. Edelman, D. R. Gandara, P. Hausner, V. Israel, D. Thornton, J. DeSanto, L. A. Doyle, Lung cancer 2003, 39, 197–199.
- [37] B. Sammet, T. Bogner, M. Nahrwold, C. Weiss, N. Sewald, J. Org. Chem. 2010, 75, 6953–6960.
- [38] C. Weiss, B. Sammet, N. Sewald, Nat. Prod. Rep. 2013, 30, 924-40.
- [39] S. Eißler, A. Stoncius, M. Nahrwold, N. Sewald, Synthesis 2006, 2006, 3747–3789.
- [40] M. Nahrwold, C. Weiß, T. Bogner, F. Mertink, J. Conradi, B. Sammet, R. Palmisano, S. Royo Gracia, T. Preuße, N. Sewald, J. Med. Chem. 2013, 56, 1853–1864.
- [41] V. A. Verma, T. H. Pillow, L. DePalatis, G. Li, G. L. Phillips, A. G. Polson, H. E. Raab, S. Spencer, B. Zheng, *Bioorg. Med. Chem. Lett.* 2015, 25, 864– 868
- [42] H. Chen, Z. Lin, K. E. Arnst, D. D. Miller, W. Li, Molecules 2017, 22, 1281.
- [43] E. Figueras, A. Borbély, M. Ismail, M. Frese, N. Sewald, *Beilstein J. Org. Chem.* 2018, 14, 1281–1286.
- [44] J. Liang, R. E. Moore, E. D. Moher, J. E. Munroe, R. S. Al-awar, D. A. Hay, D. L. Varie, T. Y. Zhang, J. A. Aikins, M. J. Martinelli, C. Shih, J. E. Ray, L. L. Gibson, V. Vasudevan, L. Polin, K. White, J. Kushner, C. Simpson, S. Pugh, T. H. Corbett, *Invest. New Drugs* 2005, 23, 213–224.
- [45] C. Steinkühler, P. Gallinari, B. Osswald, N. Sewald, M. Ritzefeld, M. Frese, E. Figueras, L. Pethö, 2016, WO2016/146638 A1.

- [46] S. Cazzamalli, E. Figueras, L. Pethő, A. Borbély, C. Steinkühler, D. Neri, N. Sewald, ACS Omega 2018, 3, 14726–14731.
- [47] A. Borbély, E. Figueras, A. Martins, S. Esposito, G. Auciello, E. Monteagudo, A. Di Marco, V. Summa, P. Cordella, R. Perego, I. Kemker, M. Frese, P. Gallinari, C. Steinkühler, N. Sewald, *Pharmaceuticals* 2019, 11, 151.
- [48] A. Dal Corso, L. Pignataro, L. Belvisi, C. Gennari, Curr. Top. Med. Chem. 2016, 16, 314–329.
- [49] D. A. Cheresh, R. C. Spiro, J. Biol. Chem. 1987, 262, 17703–17711.
- [50] N. Jain, S. W. Smith, S. Ghone, B. Tomczuk, *Pharm. Res.* 2015, 32, 3526–3540.
- [51] S. L. Goodman, H. J. Grote, C. Wilm, Biol. Open 2012, 1, 329-40.
- [52] R. J. D. Hatley, S. J. F. Macdonald, R. J. Slack, J. Le, S. B. Ludbrook, P. T. Lukey, Angew. Chem. Int. Ed. 2018, 57, 3298–3321; Angew. Chem. 2018, 130, 3354–3379.
- [53] B. Felding-Habermann, B. M. Mueller, C. A. Romerdahl, D. A. Cheresh, J. Clin. Invest. 1992, 89, 2018–2022.
- [54] B. Felding-Habermann, E. Fransvea, T. E. O'Toole, L. Manzuk, B. Faha, M. Hensler, Clin. Exp. Metastasis 2002, 19, 427–436.
- [55] J. R. Lange, W. H. Goldmann, J. L. Alonso, Biochem. Biophys. Res. Commun. 2016, 478, 1280–1285.
- [56] I. Guzzetti, M. Civera, F. Vasile, D. Arosio, C. Tringali, U. Piarulli, C. Gennari, L. Pignataro, L. Belvisi, D. Potenza, ChemistryOpen 2017, 6, 128–136.
- [57] S. Castel, R. Pagan, F. Mitjans, J. Piulats, S. Goodman, A. Jonczyk, F. Huber, S. Vilaró, M. Reina, *Lab. Invest.* 2001, 81, 1615–1626.
- [58] M. Srinivasarao, C. V. Galliford, P. S. Low, Nat. Rev. Drug Discovery 2015, 14, 203–219.
- [59] B. D. C. Chen, A. N. Nakeff, F. V. Valeriote, Int. J. Cancer 1998, 77, 869–873.

Manuscript received: March 27, 2019 Revised manuscript received: April 16, 2019